

MUNI RECETOX

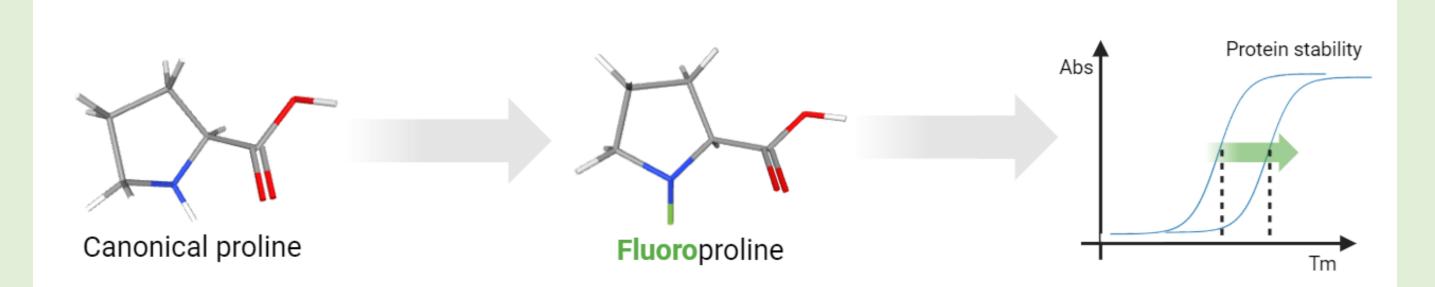
Exploring Non-Canonical Amino Acid Integration in Fibroblast Growth Factors

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BACKGROUND

Fibroblast Growth Factors (FGFs) regulate vital mechanisms in complex organisms, including cell migration, proliferation, differentiation, and survival. Their potential for *wound healing, cancer treatment*, and *tissue repair* has attracted significant interest.

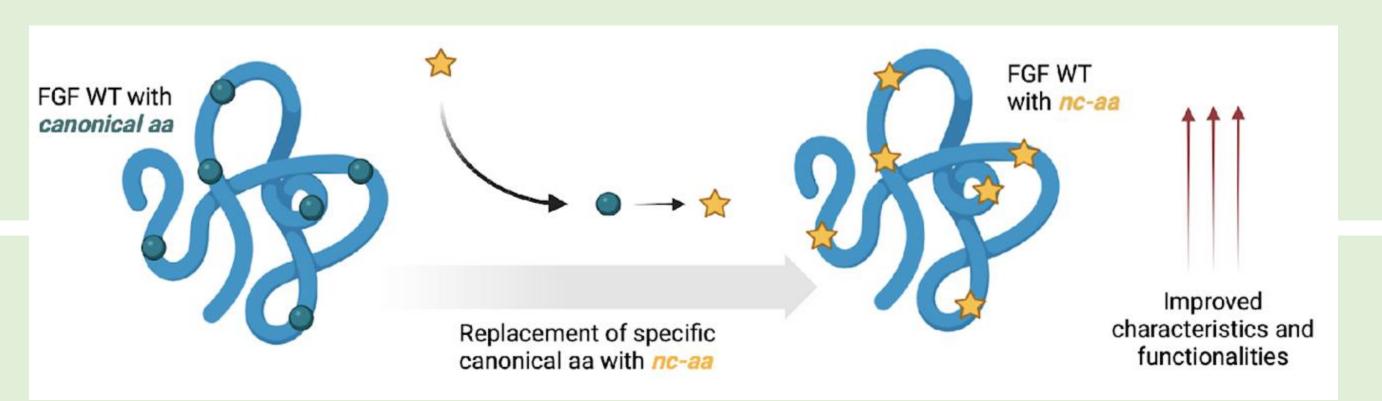
Expanding the genetic code enables the use of *non-canonical amino* acids (ncAAs), absent in the human genome. Robust methods for genetic code expansion facilitate the incorporation of ncAAs into protein polypeptide chains. This enhances *stability* and *activity* by introducing *novel functionalities* and *chemical properties* into enzymes [1].





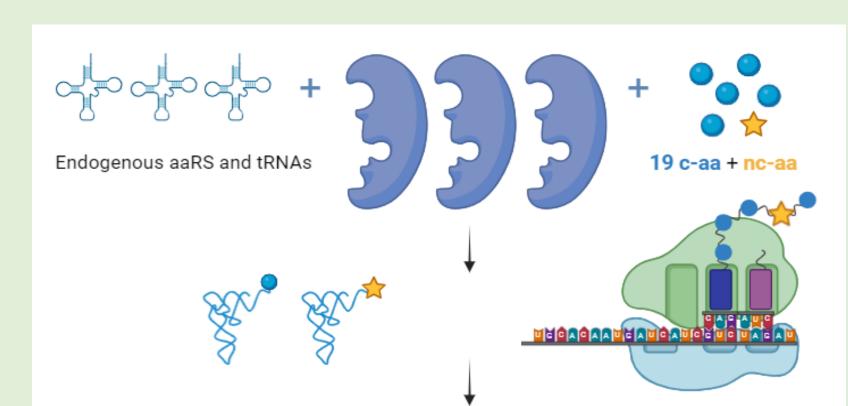
Using the **SPI method**, we aim to assess how the insertion of ncAAs affect the *stability* and *activity* of FGFs proteins and to understand the *general applicability* of nc-aa incorporation to improve FGF proteins.

In the SCS method, we introduce ncAAs at specific positions within the protein. These ncAAs serve as handles for click reactions, enabling the *immobilization* or *interaction* of FGFs with other *biological entities*.



METHODS

Two state-of-the-art methods will be used: Selective Pressure Incorporation (SPI) and Stop Codon Suppression (SCS).



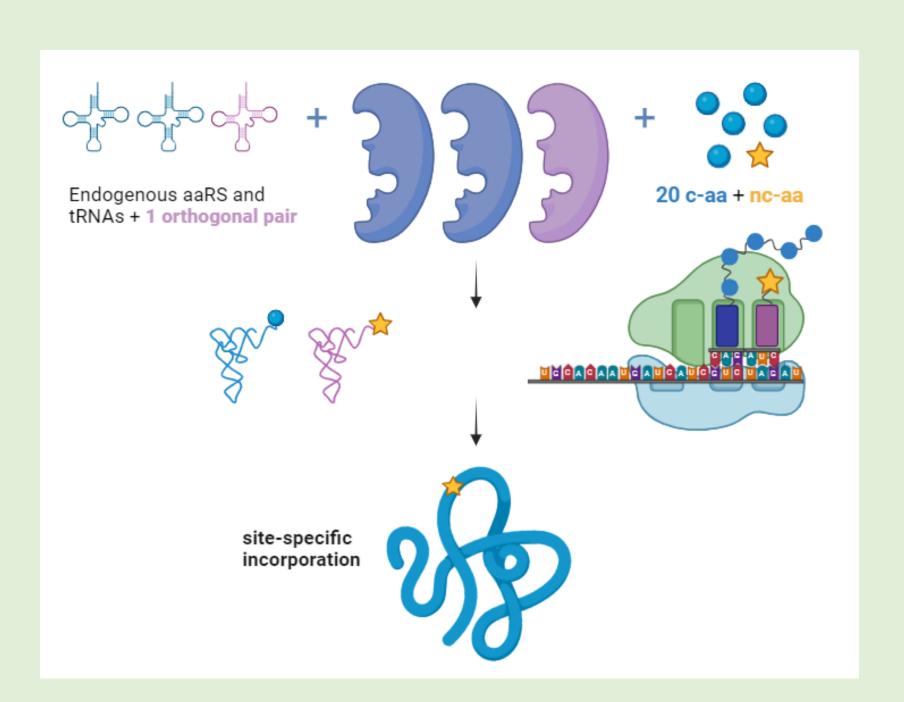
SPI

Global replacement of cAAs with the corresponding ncAAs with *residue-specific incorporation*. [2]

- It requires an *auxotrophic system* to replace the cAAs;
- It employs *native expression systems* readily available in the organism;

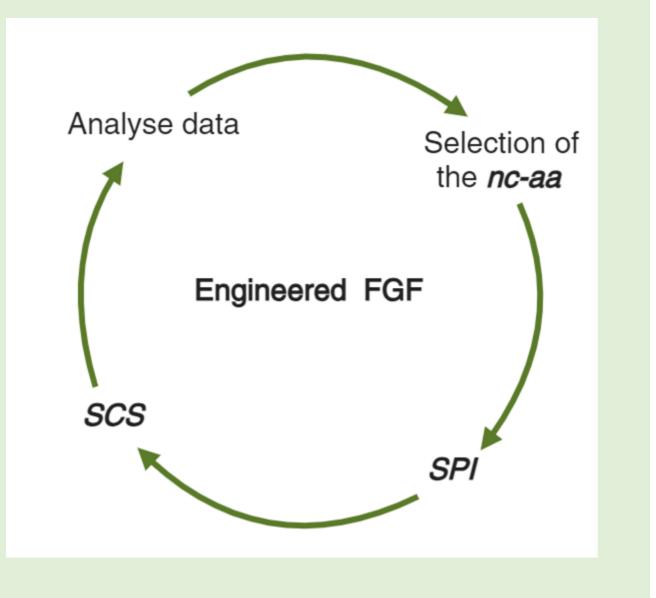


- It allows the simultaneous incorporation of multiple ncAAs into the same protein;
- SPI *does not increase* the size of the *amino acid alphabet*: cAA \rightarrow ncAA; \bullet
- It uses a pool of ncAA similar to the 20 cAA.



WORKFLOW

• Selection of the FGFs of interest along with the *appropriate nc-aa*;



SCS

Replacement of a specific natural amino acid with a nc-aa through *site-specific incorporation.* [2]

- It does *not require* an *auxotrophic system* to replace the cAAs;
- SCS uses *non-native expression systems*;
- SCS the ncAAs are added to the 20 cAA repertoire;
- SCS allows the site-specific incorporation of ncAAs at virtually any position;
- SCS allows the incorporation of a variety of surrogates distinctly different from the 20 cAAs.
- **SPI** approach to provide a general overview of the critical sites and amino acids within FGF structures;
- SCS approach to the replacement of specific amino acids for *immobilisation* or *functionalisation*;
- Analysis of collected data to understand *future implications* for industrial applications and directions to optimize FGFs with nc-aas.

CURRENT STATE

After completing a training period, I started my doctoral project with the SPI method. I decided to start with 4-fluoroproline and difluoroproline and an E. coli auxotroph strain. Following this, I made the cells competent, enabling them to efficiently uptake and express FGF2, FGF4, and FGF8 plasmids, all of interest to our company. [3]

Currently, my focus lies on optimizing the SPI technique to ensure the successful integration of non-canonical amino acids within the cells, a crucial step for enhancing FGF production.

In June, I begin a four-month research visit to Professor Budisa's lab in Canada. His expertise in the field is renowned. I aim to learn new techniques to advance my work at Enantis

REFERENCES

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- 2. Agostini, Federica, et al. "Biocatalysis with Unnatural Amino Acids: Enzymology Meets Xenobiology." Angewandte Chemie International Edition, vol. 56, no. 33, 17 July 2017, pp. 9680-9703.
- 3. Kubyshkin, V, Davis, R & Budisa, N 2021, 'Biochemistry of fluoroprolines: the prospect of making fluorine a bioelement', Beilstein Journal of Organic Chemistry, vol. 17, pp. 439-460.

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